

THE ONCOLYTIC POTENTIAL OF TARGETING THE KERATINOCYTE GROWTH FACTOR (KGF)/KGF RECEPTOR PATHWAY

J.T. Pento

Department of Pharmaceutical Sciences, College of Pharmacy, University of Oklahoma, Health Sciences Center, Oklahoma City, Oklahoma 73117, USA Correspondence: tom-pento@ouhsc.edu

CONTENTS

Abstract	915
Introduction	915
Targets	917
Conclusions	919
References	919

ABSTRACT

Growth factors and cytokines are often directly involved in cancer cell invasion and the cascade of metastatic cancer progression. These growth factors are often produced by stromal tissue, surrounding the primary tumors where they enhance cancer cell proliferation and stimulate progression to a metastatic phenotype. Keratinocyte growth factor (KGF), a member of the fibroblast growth factor (FGF) family, is known to be produced by stromal tissue and to be associated with the growth and progression of many types of cancer. KGF binds to a specific tyrosine kinase receptor which mediates the KGF cellular response via the Erk signaling pathway. Keratinocyte growth factor responsive cancer cells often contain upregulated KGF receptor (KGFR). The KGF/KGFR signaling pathway appears to be an early signal in the progression of many different types of cancer. Thus, selective inhibitors of the KGF/KGFR signaling pathway have the potential to be effective therapeutic agents for the prevention of metastatic cancer development. Furthermore, elements of this signaling pathway may be important biomarkers of cancer progression and/or therapeutic staging.

INTRODUCTION

Cancer metastasis is responsible for most of the morbidity and mortality associated with cancer. Cancer metastatic progression is a multiple-step process which includes enhanced cell proliferation, release of proteolytic enzymes, cell motility and invasion, angiogenesis and establishment of a supportive microenvironment at the sites of metastatic growth (1, 2). The molecular mechanisms that control metastasis are related to alterations in various oncogenes, tumor suppressor genes, metastasis suppressor genes and growth factors and their receptors (3).

Cell motility is another important aspect of the metastatic process which is associated with penetration of cancer cells through the

basal lamina and endothelial lining of capillaries and the entry of tumor cells into the blood and lymph vascular system. Entry of the cancer cells into the circulation is responsible for widespread dissemination of highly motile cells throughout the body. Peptide growth factors are known to be involved in cell motility and have a profound effect on metastatic progression (4). For example, the overexpression of epidermal growth factor (EGF), insulin-like growth factor, transforming growth factor and vasoactive epithelial growth factor are clearly involved in progression and metastasis of various cancers (5-9). Accordingly, keratinocyte growth factor (KGF) and/or the KGF receptor (KGFR) have been linked to the invasiveness and progression of many types of cancer. This review focuses on the potential to target the elements of the KGF/KGFR signaling pathway for the diagnosis, treatment or prevention of metastatic cancer progression.

Keratinocyte growth factor

Keratinocyte growth factor (KGF) was first observed in 1989 in a study to identify substances produced in tissue stroma that stimulate the proliferation and motility of epithelial cancer cells (10, 11). This growth factor is a protein comprising 194 amino acids which undergoes post-translational glycosylation which may not be necessary for its full biological activity (10). Originally identified in human embryonic lung fibroblasts, KGF is a member of the fibroblast growth factor (FGF) family and has also been designated FGF-7 (11). The carboxy-terminal region of KGF is 30-40% identical to other members of the FGF family (12, 13). At present, 18 FGFs (FGF1-FGF10 and FGF16-FGF 23) have been identified and grouped into six sub-families (14). Other FGF homologs (FGF11-FGF14) do not bind to or activate the known FGF receptors and thus are not currently considered part of the FGF family (15).

Reproductive hormones are directly involved in the regulation of KGF expression in reproductive stromal tissue (16-19). For example, in human endometrial tissue, the expression of KGF is progesterone-dependent while the expression of KGFR in epithelial cells is estrogen-dependent (20). Koji et al. reported that KGF transcripts in endometrial tissue of rhesus monkeys were elevated 70- to 100-fold in response to a combination of estradiol and progesterone (18). More recently, Pedchenko and Imagawa reported that prolactin and

progesterone have a direct regulatory role on KGF-induced proliferation of mouse mammary epithelium (21). Furthermore, KGF appears to play a role in development of reproductive tissue (22, 23).

Keratinocyte growth factor produced by stromal tissue surrounding the epithelial tumor tissue has been demonstrated to stimulate DNA synthesis, proliferation and migration of epithelial cancer cells (23, 24). In addition to embryonic fibroblasts, KGF has also been identified in stromal tissue from human adult lungs, skin, stomach, kidney, bladder, prostate and mammary tissue (10). Keratinocyte growth factor appears to function as a paracrine mediator of mesenchymal-epithelial interaction in embryonic development, and perhaps, wound healing homeostasis in adult tissue. Evidence of tissue regeneration and wound healing has been demonstrated in cornea, skin, lung and tympanic membrane tissue and hair follicles (15, 23, 25, 26). Furthermore, tissue KGF levels have been shown to increase dramatically following skin, bladder and kidney injury (27-29).

Keratinocyte growth factor also appears to enhance tissue cytoprotection in oral mucosa and tissue along the gastrointestinal tract. These observations have led to the development of palifermin, which is an *N*-terminally truncated derivative of KGF with increased stability, as a cytoprotective agent to prevent or reduce the severity of radiation- or chemotherapy-induced mucositis in bone marrow transplant patients (30). Palifermin has been demonstrated to increase epithelial cell proliferation and the thickness of the oral epithelium for a week (31-33). Furthermore, palifermin and repifermin (a similar FGF10 peptide) are under investigation to improve wound healing, immune function and to treat ulcerative colitis (34-37).

The biological actions of KGF are thought to be involved in normal morphogenesis and tissue repair; however, KGF may contribute to tumor cell progression by enhancing cancer cell proliferation, motility and invasion (26, 38-41). Furthermore, it appears that upregulation or overexpression of KGF and/or KGFR is associated with cancer progression and metastatic development (10, 42). Accordingly, KGF, KGFR and intermediates in the signal transduction pathways may represent valuable therapeutic targets to reduce or prevent metastatic development.

Keratinocyte growth factor receptor

Keratinocyte growth factor receptor (KGFR, also referred to as FGFR2IIIb) is a splice variant of FGFR-2 which is encoded by the *FGFR-2* gene (43, 44). KGFR is a member of the fibroblast growth factor receptor (FGFR) family consisting of four known peptides (FGFR1-FGFR4). The FGF receptors are all membrane-spanning tyrosine kinase receptors with highly conserved amino acid sequences (39). These receptors consist of 3 extracellular immunoglobulin domains, a transmembrane domain and a cytoplasmic tyrosine kinase domain (45). Alternate splicing of the extracellular domain results in several isoforms with distinct binding specificity (46). It is well established that the target epithelial cells contain high-affinity KGFRs (12, 26, 44). In situ hybridization studies confirmed the specific mesenchymal distribution of KGF and the epithelial distribution of KGFR in target tissue. This observation provides further evidence that KGF is a mesenchymally derived mediator of epithelial cell proliferation and migration (10, 47). As with KGF, the KGFR is associated with normal tissue morphogenesis and repair (37, 38, 48-50). However, KGFR is known to be overexpressed in

many types of types of cancer and changes in its binding specificity are often associated with cancer progression (51). Accordingly, Jang and coworkers reported gain-of-function mutations with KGFR in gastric and colorectal cancer (52).

KGF/KGFR pathway signaling

Keratinocyte growth factor/KGFR signaling is known to involve either the extracellular signal-regulated kinases 1,2 (Erk 1,2) pathway or the 3-phosphoinositide-dependent protein kinase (PI3K) pathway (25, 53-55). Apparently the primary signaling pathway which mediates KGF-induced cancer cell motility involves the Erk 1,2 kinase pathway (55, 56). Accordingly, the Erk 1,2 signaling pathway mediates both the cell proliferation and cell motility associated with KGF-mediated wound healing (57).

An analysis of gene expression in MCF-7 breast cancer cells revealed that KGF treatment produces an increase in the mRNA levels of growth factor receptor bound protein-2 (Grb2) and other Erk signaling intermediates (58). The *Grb2* gene is very highly conserved among species. Moreover, the Grb2 protein is a ubiquitously expressed adaptor protein. Growth factor receptor bound protein-2 is also known to activate tyrosine kinase signaling via Ras (59), which could in turn activate the Raf/MEK/Erk1,2 pathway (60). Tari et al. demonstrated that downregulation of Grb2 protein expression by with Grb2 antisense oligonucleotides can inhibit the proliferation of breast cancer cells in an Erk 1,2-dependent manner (61). Grb2 is also known to be associated with FGF receptors (62) and has been shown to be involved in cell motility (63). The Grb2 protein was observed to be overexpressed in breast cancer cells and in specimen breast cancer tissue (64, 65). Furthermore, it has been observed that KGF treatment doubles the expression of phospho-Erk 1,2, while downregulation of Grb2 expression inhibits KGF-mediated breast cancer cell motility (56). Accordingly, Erk 1,2 activity is known to be involved in the proliferation of endometrial carcinoma cells and associated with KGF-mediated invasiveness of stomach cancer cells (53, 55).

It has also been suggested that signal transduction associated with the FGFR involves the binding of heparin sulfate, an abundant cell surface molecule, to form a ternary signaling complex (66). It has been further suggested that heparin proteoglycans on the surface of cancer cells may attract KGF to cell surface receptors. In addition, the proteoglycans may serve as a reservoir for KGF on the cell membrane, thus making this growth factor available for KGFR activation and enhanced signaling (67).

Involvement of the KGF/KGFR pathway in cancer

Enhanced cancer cell motility is usually observed during the metastatic dissemination of tumor cells to secondary sites (68). This stimulation of motile cell behavior is known to be regulated by specific cytokines and growth factors (69, 70). In many cases, the stromal tissue which surrounds the primary tumor cells produces growth factors, which stimulate tumor cell motility and proliferation; hence progression to a more metastatic phenotype (71, 72).

Recently, Zang et al. used a cancer cDNA profiling assay to examine the expression of KGFR in 154 tumor samples and paired normal samples representing 19 types of human cancer (73). The researchers observed that KGFR was upregulated in many reproduc-

tive and other types of cancer tissue at an early stage of cancer development (i.e., uterus, cervix, vulva, prostate, testes and lung). These results support the concept that that KGFR upregulation may be an early event in the progression of these cancers and that KGFR expression levels may be a useful prognostic biomarker. However, it was found that KGFR was upregulated in more advanced tumors in other types of cancer (i.e., ovary, stomach, small intestine, rectum, bladder, trachea and pancreas), while it was downregulated in some other types of cancer on the array (i.e., skin, liver, colon and kidney). As highlighted in the various examples above, KGF signaling appears to be involved in the progression of many types of cancer and, therefore, has the potential to serve as a useful oncolytic biomarker.

A summary of the literature on the involvement of the KGF/KGFR pathway in various common cancers follows.

TARGETS

Breast cancer

Mammary glands of adult female animals are remarkably sensitive to KGF (74). For example, systemic administration of KGF in nulliparous female rats for 3 days was found to produce massive mammary ductal hyperplasia and an elevation of mitotic figures (74). Similarly, administration of KGF to adult male and pregnant female rats produced ductal hyperplasia and acinar proliferation. Accordingly, intraductal hyperplasia is well known to be characteristic of premalignant breast lesions which leads to neoplasia (10). Kitsberg and Leder created a strain of transgenic mice that carry a constitutively upregulated KGF transgene (75). They observed that female mice with this transgene develop very dramatic mammary epithelial hyperplasia and eventually all animals developed metastatic mammary carcinomas. Consistent with these results, KGFR gene upregulation was observed in human primary breast tumor samples (76). It was reported that expression of KGFR occurs in malignant and non-malignant breast cancer cell lines; however, it was also observed that highly malignant, metastatic breast cancer tissue expressed relatively little KGFR (77). These findings suggest that KGF-mediated stimulation of breast epithelial proliferation and migration may be an early event in the molecular cascade that leads to breast cancer progression and metastasis (78). Thus, KGF may stimulate the motility of well-differentiated breast cancer cells (e.g., ER positive breast cancer) and have little or no effect on less differentiated, highly malignant tumor cells that have escaped normal regulatory mechanisms. Furthermore, KGF may not stimulate highly malignant tumor cells because KGFR-associated signal transduction is constitutively upregulated, and thus these cells would be unresponsive to additional KGF stimulation. In agreement with these statements, Zang et al. observed that treatment of ER-positive human breast cancer cells with recombinant human KGF produced a profound stimulation of cell motility and an upregulation of the *KGFR* gene. Researchers also found that this effect did not occur in ER-negative cell lines (79). In another study, KGF motility response in breast cancer cells proved to be dose-dependent and characterized by an immediate increase in ruffling of the plasma membrane and cell scattering which continued for up to 48 hours following KGF treatment (80). Furthermore, transfection of MCF-7 cells with a KGF-producing vector was observed to enhance cell motility and to produce

a characteristic motile morphology (81). Such changes in cell morphology, associated with membrane ruffling and motility, are believed to be associated with cytoskeletal reorganization and necessary for adhesion foci, cell surface ligand-receptor binding and regulation of gene transduction (82, 83). Accordingly, a significant alteration in the distribution of f-actin, an indicator of cytoskeletal reorganization, was observed in the cytoplasm of KGF-stimulated MCF-7 cells (84).

Both KGF and KGFR have been reported to enhance the progression of breast cancer by inhibiting normal apoptosis via the overexpression of Bcl-2 (85). Furthermore, results of a recent study indicate that KGF-mediated signaling involves the *Wilms' Tumor 1 (WT1)* and *focal adhesion kinase (FAK)* genes (86). Interestingly, both *WT1* and *FAK* are involved in the regulation of Grb2 and signaling via the Erk pathway (87-89).

Taken together these observations suggest that KGF-mediated stimulation of breast epithelial proliferation and migration may be an important step in breast cancer progression and metastasis (90). Furthermore, KGF expression correlates closely with estrogen receptor (ER)- α expression in human breast cancer tissue, and the promoter region of the *KGF* gene is known to contain a semi-palindromic sequence of the estrogen response element (91). It has been demonstrated that selective inhibition of ER- α using a selective RNAi reduced KGF-mediated stimulation of breast cancer cells (92, 93). These findings illustrate the potential value of KGF/KGFR expression levels to serve as biomarkers for breast cancer progression and/or responsiveness to chemotherapy.

Prostate cancer

The expression of both KGF and KGFR were found to be elevated in benign prostatic hypertrophy (BPH) and in prostate cancer where KGF appears to act in a paracrine manner in cancer progression (94-96). It has been proposed that KGF acts together with androgens in the stimulation of cell proliferation in BPH (97, 98). Using immunolocalization techniques, Planaz and coworkers detected KGF in epithelial cells from both BPH and prostate cancer tissue and suggested that KGF stimulates cell proliferation in an autocrine manner during cancer development (99). In clinical prostate cancer the expression of KGF and KGFR increases with disease progression. Furthermore, the switching from stromal to epithelial regulation, which permits the cancer cells to escape normal stromal regulation, occurs during metastatic progression (100, 101). Accordingly, switching of the KGFR from the FGFR2-IIIb (androgen-regulated receptor isoform) to the FGFR2-IIIc isoform, which recognizes other FGF peptides, occurs during tumor progression and enhances unregulated tumor growth and invasion (71, 100, 102-104). Thus, monitoring these receptor isoforms may serve as an early biomarker for progression of prostate cancer (105).

Colon cancer

Overexpression of KGFR has been observed in colorectal cancer cells (106) and KGF was found to produce a dose-dependent stimulation of well-differentiated colorectal cancer cells, but not metastatic cancer cells or poorly differentiated colorectal cancer cells (107, 108). Yoshino and coworkers reported that KGFR is overexpressed in

tumor samples from approximately two-thirds of colorectal cancer patients (109). Since the receptor was localized at the center of cancer nests and co-localized with cytokeratin-20, which is known to be a diagnostic marker for colorectal cancer, this co-localization of both may improve the predictive therapeutic accuracy of these biomarkers (110). Keratinocyte growth factor receptor expression was associated with a well-differentiated histology, suggesting that KGFR signaling occurs early in the development of colorectal cancer (109). Wantanabe and coworkers who examined KGF and KGFR expression in colorectal cancer tissue from 12 patients, proposed that KGF acts in a paracrine and autocrine manner to induce cell growth in this cancer (111).

Lung cancer

Keratinocyte growth factor is involved in embryonic lung development and in the adult lung it appears to be involved in the homeostasis and repair of alveolar and bronchial epithelial cells (50, 74, 112). However, in a study of KGF and KGFR expression in tissue from human lung cancer patients, Yamayoshi and coworkers (113) observed that co-expression of KGF and KGFR was associated with greater differentiation in squamous cell carcinomas, while in lung carcinomas KGF co-expression was associated with poor differentiation, metastatic involvement and a shorter duration of patient survival. These investigators speculated that KGF/KGFR expression and signaling may be useful biomarkers for the grading of lung carcinoma. Furthermore, Baumann et al. have proposed the use of KGF as a molecular target for radiotherapy of lung cancer (114).

Ovarian, uterine and cervical cancers

Steele and coworkers reported that KGFR is undetectable in normal ovarian epithelial cells, while 80% of the ovarian cancer samples expressed KGFR (115). It was suggested that KGFR overexpression is involved in the progression of ovarian cancer by making the premalignant tissue more responsive to paracrine KGF stimulation (115, 116). This argument is supported by research by Parrott et al., who reported that KGF produced in the ovarian tumor microenvironment may be involved in the progression of this cancer (117). Similarly, KGFR was detected in carcinomas from 86% of cervical cancer patients, suggesting that KGFR mediates the progression of reserve and squamous metaplastic cells in cervical cancer tissue (118). Furthermore, several groups have reported that the KGF signaling pathway enhances the proliferation of human endometrial cancer and may be a useful progression biomarker (55, 119). Accordingly, KGF expression appears to be regulated by estradiol, progesterone and gonadotropins (hormones that are known to regulate the normal growth and cycling of ovarian, uterine and cervical tissues; 19, 117).

Pancreatic cancer

Carcinoma of the pancreas is one of the most deadly forms of cancer, with a death rate approaching the incidence rate, largely because most patients have advanced, metastatic disease at the time of diagnosis (120). Keratinocyte growth factor expression and KGFR signaling appear to be associated with enhanced proliferation and the progression of human pancreatic cancer. For example, Siddiqi and coworkers observed that KGF expression was enhanced in 7 of 16 human pancreatic cancer samples and that 5 of 7 pancreatic

cancer cell lines expressed KGFR (121). Similarly, overexpression of KGFR in 7 of 10 pancreatic cancer samples was observed using a cDNA cancer profiling array comparing tumor and patient matched normal tissue (73). Interestingly, it was reported that KGF and KGFR expression levels were increased 5-fold within 28 days in a rat model of pancreatitis in vivo (122). In addition, overexpression and co-localization of KGF and KGFR in pancreatic cancer and adjacent parenchyma has been observed, indicating that KGF may enhance the progression of pancreatic cancer in either an autocrine or paracrine manner (123). Similarly, during disease progression other FGFs and FGF receptors are overexpressed and appear to be involved in the invasiveness of pancreatic cancer (124-126). In a recent study of human pancreatic cancer cells it was reported that KGF treatment significantly enhanced the proliferation and motility of the pancreatic cancer cells within a period of 24-48 hours (127).

The above research suggest that upregulation of KGF secretion and/or KGFR overexpression may be important biomarkers and, therefore, signals in the progression of pancreatic cancer (121, 123, 128). Consequently, inhibition of KGFR signaling and/or inhibition of genes and proteins regulated by KGF/KGFR signaling may impede the progression of pancreatic cancer cells to a more metastatic phenotype (126, 129).

Stomach cancer

As early as 1990, researchers observed that *KATO-III cell-derived stomach cancer amplified* gene, which is 88% homologous to *KGFR*, was upregulated in poorly differentiated stomach cancer tissue (130). An upregulation of the *KGFR* gene of as much as 30- to 40-fold in gastric cancer compared to normal tissue has also been reported (73, 131). Keratinocyte growth factor from gastric fibroblasts or added to the culture media was found to stimulate the proliferation of scirrhous gastric carcinoma cell lines (132). Furthermore, KGF treatment enhanced the production of matrix metalloprotease-9 and urokinasetype plasminogen activator (both associated with cancer cell invasion) in these human stomach cancer cells (53). Matsunobu and coworkers reported that KGFR expression was associated with expansive growth and shallow gastric wall invasion at an early stage of stomach cancer (133). These studies suggest that KGFR expression may be used as a tool to identify stomach cancer progression.

Targeting the KGF/KGFR pathway

It appears that an upregulation of KGF/KGFR signaling at an early stage of cancer development is involved in the progression of a number of common cancers as described above and as a result, selective inhibitors of this signaling pathway may be effective oncolytic therapeutic agents.

KGF antagonists

Fibroblast growth factors are a family of peptides which share unique binding affinity for heparin. Accordingly, it appears that binding to heparin and other cell surface proteoglycans enhances the activity of these FGF peptides (134). Heparin proteoglycans on the cell surface may act to bind the FGF growth factors, increasing their availability to cell surface receptors and serving as a reservoir for the

FGF growth factors on the cell membrane (135). However, heparin is reported to inhibit the action of KGF, unlike other members of the FGF family (135, 136).

It has been known for decades that anticoagulant treatment with heparin reduces the growth and metastatic development of various cancers (134). Retrospective meta-analysis of clinical studies for the prevention of venous thromboembolism, employing heparin and/or low-molecular-weight heparin (LMWH), has indicated that their therapeutic use is associated with a reduction in cancer-associated mortality (137). Low-molecular-weight heparin is more stable and has better pharmacokinetic properties than heparin and, accordingly, most of these studies found greater anticancer activity with LMWH therapy (134, 138). The mechanism for this oncolytic effect has not been established, but it has been suggested that inhibition of FGF-mediated tumor angiogenesis may be responsible for this activity (26, 134). However, in light of the fact that heparin inhibits KGF, the reduction in KGF activity may also be involved in the beneficial effects of heparin treatment in improving cancer patient survival.

The KGFR2 β (IIIb)/Fc chimera is a soluble extracellular fragment of KGFR which acts as a highly selective KGF inhibitor (139). Accordingly, KGF binds to this KGFR fragment with much greater selectivity than to the heparin-related compounds (140). The results of a study which compared the influence of heparin, LMWH and KGFR2 β (IIIb)/Fc on KGF induced breast cancer cell motility and proliferation demonstrated that all three were equally effective KGF antagonists during the first several hours of treatment. However, after 2 hours the heparin-mediated inhibition of KGF activity diminished, while LMWH and KGFR2 β (IIIb)/Fc produced a more prolonged inhibitory effect lasting up to 48 hours (139).

In addition, there are a number of other heparin-like compounds or heparinoids which may be useful in the treatment of cancer (141-143). Suramin, a polysulfated naphthylurea which produces heparin-like inhibition of FGF signaling, has been shown to be effective in the treatment of bladder, kidney and prostate cancers (144-146). Furthermore, suramin treatment may enhance the effectiveness of other chemotherapeutic agents in prostate cancer (147). Similarly, Rotolo et al. reported that silencing the expression of KGFR restored the responsiveness of breast cancer cells to 5-fluorouracil and speculated that selective inhibition of the KGF/KGFR pathway may prevent resistance to chemotherapy (148). Other promising heparinoids include PI-88, which acts as a heparinase inhibitor (149), and BXL-628, a vitamin D analogue, which is a KGF inhibitor that has been demonstrated to inhibit the proliferation and invasiveness of prostate cancer cells (150).

These compounds have the potential to be useful in the treatment or prevention of cancer progression associated with KGF signaling. However, the anticoagulant activity and poor oral bioavailability for most heparin-related compounds may limit their long-term use in cancer therapy.

KGFR tyrosine kinase antagonists

Changes in the regulation of growth factor receptor tyrosine kinase (TK) activity and the related signal transduction is known to be involved in the development and progression cancer (66, 151, 152).

As an example, it is well known that EGF receptor overexpression is predictive of aggressive and metastatic cancer development (153-155) and selective EGF receptor TK inhibitors have been demonstrated to be therapeutically effective in the treatment of various cancers (153, 156, 157).

In 1993, Yan and coworkers established that a transition in the KGFR occurs from the initial (FGFR IIIb) isoform found in primary tumors, which is KGF responsive, to the FGFR IIIc isoform found in more advanced cancer, which is unresponsive to KGF (100). This transition of KGFR isoforms during cancer progression suggests that KGFR activation is an important early step in the initiation metastatic cancer progression. Therefore, selective inhibition of the FGFR-IIIb isoform would provide an opportunity to prevent or reduce cancer progression to a more malignant phenotype.

Nonselective inhibitors of mitogen-activated protein kinase signaling have been demonstrated to inhibit KGF stimulation of breast and endometrial cancer cells (55, 56, 158). These nonselective inhibitors alter the signaling activity of numerous growth factors which would likely result in unacceptable adverse side effects. However, selective KGFR TK antagonists may be relatively nontoxic and therapeutically effective oncolytic agents. Since KGFR is a growth factor TK receptor; the design of small-molecule, selective inhibitors of KGFR that compete with adenosine triphosphate (ATP) for the catalytic site in the receptor is a viable approach that has been employed (159, 160).

In efforts to design novel KGFR-selective ATP site-directed ligands, Hackett et al. employed in silico site-directed mutagenesis to generate a homology model of the KGFR TK domain. The compounds identified with this modeling algorithm were found to inhibit KGF-mediated breast cancer cell proliferation and motility in a culture wounding model. Furthermore, the researchers observed that the most potent KGF inhibitor produced a reduction in the density of KGFR on breast cancer cells, suggesting that receptor targeting downregulates the expression of KGFR (161). Therefore, these selective KGFR TK antagonists may be useful therapeutically to target the KGFR signaling pathway and prevent or significantly reduce cancer progression.

CONCLUSIONS

It is clear that the progression of cancer to a metastatic phenotype is largely responsible for cancer mortality. Today very few therapeutic modalities with the ability to selectively inhibit or reduce metastatic progression are available. We know that upregulation of KGF and KGFR appears to be involved in cancer cell proliferation, motility and enhanced survival. Because KGF/KGFR signaling appears to be associated with the progression of many types of cancer at an early stage, the measurement of tissue or circulating levels of KGF/KGFR or related signaling may serve as important biomarkers of cancer progression and/or therapeutic monitoring. Moreover, agents which are selective and potent inhibitors of the KGF/KGFR signaling pathway may have considerable oncolytic potential.

REFERENCES

1. Poste, G., Fidler, I.J. *The pathogenesis of cancer metastasis*. *Nature* 1980, 283(5743): 139-46.
2. Kohn, E.C., Liotta, L.A. *Molecular insights into cancer invasion: Strategies for prevention and intervention*. *Cancer Res* 1995, 55(9): 1856-62.

3. Devarajan, E., Huang, S. *STAT3 as a central regulator of tumor metastases*. *Curr Mol Med* 2009, 9(5): 626-33.
4. Gherardi, E., Stoker, M. *Hepatocytes and scatter factor*. *Nature* 1990, 346(6281): 228.
5. Welm, A.L. *TGFbeta primes breast tumor cells for metastasis*. *Cell* 2008,133(1): 27-8.
6. Kim, H., Muller, W.J. *The role of the epidermal growth factor receptor family in mammary tumorigenesis and metastasis*. *Exp Cell Res* 1999, 253(1): 78-87.
7. Romano, G. *The complex biology of the receptor for the insulin-like growth factor-I*. *Drug News Perspect* 2003, 16(8): 525-31.
8. Dvorak, H.F. *Vascular permeability factor/vascular endothelial growth factor: A critical cytokine in tumor angiogenesis and a potential target for diagnosis and therapy*. *J Clin Oncol* 2002, 20(21): 4368-80.
9. Wells, A. *Tumor invasion: Role of growth factor-induced cell motility*. *Adv Cancer Res* 2000, 78:31-101.
10. Rubin, J.S., Bottaro, D.P., Chedid, M., Mikki, T., Ron, D., Cunha, G., Finch, P.W. *Keratinocyte growth factor as a cytokine that mediates mesenchymal-epithelial interaction*. In: *Epithelial-Mesenchymal Interactions in Cancer*. Goldberg, I.D., Rosen, E.M. (Eds). Birkhäuser Verlag, Basel, Switzerland 1995, 191-214.
11. Rubin, J.S., Osada, H., Finch, P.W., Taylor, W.G., Rudikoff, S., Aaronson, S.A. *Purification and characterization of a newly identified growth factor specific for epithelial cells*. *Proc Natl Acad Sci U S A* 1989, 86(3): 802-6.
12. Aaronson, S.A., Bottaro, D.P., Miki, T. et al. *Keratinocyte growth factor. A fibroblast growth factor family member with unusual target cell specificity*. *Ann N Y Acad Sci* 1991, 638: 62-77.
13. [No author listed]. *The fibroblast growth factor family. Nomenclature meeting report and recommendations. January 17, 1991*. *Annual NY Academy of Science* 1991, 638: xiii-xvi.
14. Itoh, N., Ornitz, D.M. *Evolution of the Fgf and Fgfr gene families*. *Trends Genet* 2004, 20(11): 563-9.
15. Beenken, A., Mohammadi, M. *The FGF family: Biology, pathophysiology and therapy*. *Nat Rev Drug Discov* 2009, 8(3): 235-53.
16. Fasciana, C., van der Made, A.C., Faber, P.W., Trapman, J. *Androgen regulation of the rat keratinocyte growth factor (KGF/FGF7) promoter*. *Biochem Biophys Res Commun* 1996, 220(3): 858-63.
17. Kim, P.J., Sakaguchi, K., Sakamoto, H. et al. *Colocalization of heparin and receptor binding sites on keratinocyte growth factor*. *Biochemistry* 1998, 37(25): 8853-62.
18. Koji, T., Chedid, M., Rubin, J.S., Slayden, O.D., Csaky, K.G., Aaronson, S.A., Brenner, R.M. *Progesterone-dependent expression of keratinocyte growth factor mRNA in stromal cells of the primate endometrium: Keratinocyte growth factor as a progestomedin*. *J Cell Biol* 1994, 125(2): 393-401.
19. Parrott, J.A., Skinner, M.K. *Developmental and hormonal regulation of keratinocyte growth factor expression and action in the ovarian follicle*. *Endocrinology* 1998, 139(1): 228-35.
20. Siegfried, S., Pekonen, F., Nyman, T., Ammälä, M., Rutanen, E.M. *Distinct patterns of expression of keratinocyte growth factor and its receptor in endometrial carcinoma*. *Cancer* 1997, 79(6): 1166-71.
21. Pedchenko, V.K., Imagawa, W.T. *Mammogenic hormones differentially modulate keratinocyte growth factor (KGF)-induced proliferation and KGF receptor expression in cultured mouse mammary gland epithelium*. *Endocrinology* 1998, 139(5): 2519-26.
22. Hirai, Y., Lochter, A., Galosy, S., Koshida, S., Niwa, S., Bissell, M.J. *Epimorphin functions as a key morphoregulator for mammary epithelial cells*. *J Cell Biol* 1998, 140(1): 159-69.
23. Rubin, J.S., Bottaro, D.P., Chedid, M. et al. *Keratinocyte growth factor*. *Cell Biol Int* 1995, 19(5): 399-411.
24. Finch, P.W., Cunha, G.R., Rubin, J.S., Wong, J., Ron, D. *Pattern of keratinocyte growth factor and keratinocyte growth factor receptor expression during mouse fetal development suggests a role in mediating morphogenetic mesenchymal-epithelial interactions*. *Dev Dyn* 1995, 203(2): 223-40.
25. Chandrasekhar, G., Kakazu, A.H., Bazan, H.E. *HGF- and KGF-induced activation of PI-3K/p70 s6 kinase pathway in corneal epithelial cells: Its relevance in wound healing*. *Exp Eye Res* 2001, 73(2): 191-202.
26. Powers, C.J., McLeskey, S.W., Wellstein, A. *Fibroblast growth factors, their receptors and signaling*. *Endocr Relat Cancer* 2000, 7(3): 165-97.
27. Baskin, L.S., Sutherland, R.S., Thomson, A.A. et al. *Growth factors in bladder wound healing*. *J Urol* 1997, 157(6): 2388-95.
28. Ichimura, T., Finch, P.W., Zhang, G., Kan, M., Stevens, J.L. *Induction of FGF-7 after kidney damage: A possible paracrine mechanism for tubule repair*. *Am J Physiol* 1996, 271(5 Pt. 2): F967-76.
29. Werner, S., Peters, K.G., Longaker, M.T., Fuller-Pace, F., Banda, M.J., Williams, L.T. *Large induction of keratinocyte growth factor expression in the dermis during wound healing*. *Proc Natl Acad Sci U S A* 1992, 89(15): 6896-900.
30. Spielberger, R., Stiff, P., Bensinger, W. et al. *Palifermin for oral mucositis after intensive therapy for hematologic cancers*. *N Engl J Med* 2004, 351(25): 2590-8.
31. Beaven, A.W., Shea, T.C. *The effect of palifermin on chemotherapy and radiation therapy-induced mucositis: A review of the current literature*. *Support Cancer Ther* 2007, 4(4): 188-97.
32. Potten, C.S., Booth, D., Cragg, N.J., O'Shea, J.A., Tudor, G.L., Booth, C. *Cell kinetic studies in the murine ventral tongue epithelium: The effects of repeated exposure to keratinocyte growth factor*. *Cell Prolif* 2002, 35 Suppl 1:22-31.
33. Potten, C.S., Booth, D., Cragg, N.J. et al. *Cell kinetic studies in the murine ventral tongue epithelium: Mucositis induced by radiation and its protection by pretreatment with keratinocyte growth factor (KGF)*. *Cell Prolif* 2002, 35 Suppl 1: 32-47.
34. Freytes, C.O., Ratanatharathorn, V., Taylor, C. et al. *Phase I/II randomized trial evaluating the safety and clinical effects of repifermin administered to reduce mucositis in patients undergoing autologous hematopoietic stem cell transplantation*. *Clin Cancer Res* 2004, 10(24): 8318-24.
35. Panoskaltsis-Mortari, A., Ingbar, D.H., Jung, P. et al. *KGF pretreatment decreases B7 and granzyme B expression and hastens repair in lungs of mice after allogeneic BMT*. *Am J Physiol Lung Cell Mol Physiol* 2000, 278(5): L988-99.
36. Sandborn, W.J., Sands, B.E., Wolf, D.C. et al. *Repifermin (keratinocyte growth factor-2) for the treatment of active ulcerative colitis: A randomized, double-blind, placebo-controlled, dose-escalation trial*. *Aliment Pharmacol Ther* 2003, 17: 1355-64.
37. Werner, S. *Keratinocyte growth factor: A unique player in epithelial repair processes*. *Cytokine Growth Factor Rev* 1998, 9(2): 153-65.
38. Alarid, E.T., Rubin, J.S., Young, P., Chedid, M., Ron, D., Aaronson, S.A., Cunha, G.R. *Keratinocyte growth factor functions in epithelial induction during seminal vesicle development*. *Proc Natl Acad Sci U S A* 1994, 91(3): 1074-8.
39. Celli, G., LaRochelle, W.J., Mackem, S., Sharp, R., Merlino, G. *Soluble dominant-negative receptor uncovers essential roles for fibroblast growth factors in multi-organ induction and patterning*. *Embo J* 1998, 17(6): 1642-55.
40. Ornitz, D.M., Xu, J., Colvin, J.S. et al. *Receptor specificity of the fibroblast growth factor family*. *J Biol Chem* 1996, 271(25): 15292-7.
41. Sugimura, Y., Foster, B.A., Hom, Y.K. et al. *Keratinocyte growth factor (KGF) can replace testosterone in the ductal branching morphogenesis of the rat ventral prostate*. *Int J Dev Biol* 1996, 40(5): 941-51.

42. Finch, P.W., Rubin, J.S. *Keratinocyte growth factor expression and activity in cancer: implications for use in patients with solid tumors*. J Natl Cancer Inst 2006, 98(12): 812-24.
43. Miki, T., Bottaro, D.P., Fleming, T.P., Smith, C.L., Burgess, W.H., Chan, A.M., Aaronson, S.A. *Determination of ligand-binding specificity by alternative splicing: two distinct growth factor receptors encoded by a single gene*. Proc Natl Acad Sci U S A 1992, 89(1): 246-250.
44. Miki, T., Fleming, T.P., Bottaro, D.P., Rubin, J.S., Ron, D., Aaronson, S.A. *Expression cDNA cloning of the KGF receptor by creation of a transforming autocrine loop*. Science 1991, 251(4989): 72-5.
45. Mohammadi, M., Olsen, S.K., Ibrahimi, O.A. *Structural basis for fibroblast growth factor receptor activation*. Cytokine Growth Factor Rev 2005, 16(2): 107-37.
46. Johnson, D.E., Lu, J., Chen, H., Werner, S., Williams, L.T. *The human fibroblast growth factor receptor genes: A common structural arrangement underlies the mechanisms for generating receptor forms that differ in their third immunoglobulin domain*. Mol Cell Biol 1991, 11(9): 4627-34.
47. Mason, I.J., Fuller-Pace, F., Smith, R., Dickson, C. *FGF-7 (keratinocyte growth factor) expression during mouse development suggests roles in myogenesis, forebrain regionalisation and epithelial-mesenchymal interactions*. Mech Dev 1994, 45(1): 15-30.
48. Jackson, D., Bresnick, J., Rosewell, I., Crafton, T., Poulson, R., Stamp, G., Dickson, C. *Fibroblast growth factor receptor signalling has a role in lobuloalveolar development of the mammary gland*. J Cell Sci 1997, 110 (Pt 11): 1261-8.
49. Cunha, G.R. *Role of mesenchymal-epithelial interactions in normal and abnormal development of the mammary gland and prostate*. Cancer 1994, 74(3 Suppl): 1030-44.
50. Post, M., Souza, P., Liu, J., Tseu, I., Wang, J., Kuliszewski, M., Tanswell, A.K. *Keratinocyte growth factor and its receptor are involved in regulating early lung branching*. Development 1996, 122(10): 3107-15.
51. Grose, R., Dickson, C. *Fibroblast growth factor signaling in tumorigenesis*. Cytokine Growth Factor Rev 2005, 16(2): 179-86.
52. Jang, J.H., Shin, K.H., Park, J.G. *Mutations in fibroblast growth factor receptor 2 and fibroblast growth factor receptor 3 genes associated with human gastric and colorectal cancers*. Cancer Res 2001, 61(9): 3541-3.
53. Shin, E.Y., Ma, E.K., Kim, C.K., Kwak, S.J., Kim, E.G. *Src/ERK but not phospholipase D is involved in keratinocyte growth factor-stimulated secretion of matrix metalloproteinase-9 and urokinase-type plasminogen activator in SNU-16 human stomach cancer cell*. J Cancer Res Clin Oncol 2002, 128(11): 596-602.
54. Ray, P., Devaux, Y., Stolz, D.B. et al. *Inducible expression of keratinocyte growth factor (KGF) in mice inhibits lung epithelial cell death induced by hyperoxia*. Proc Natl Acad Sci U S A 2003, 100(10): 6098-103.
55. Taniguchi, F., Harada, T., Sakamoto, Y., Yamauchi, N., Yoshida, S., Iwabe, T., Terakawa, N. *Activation of mitogen-activated protein kinase pathway by keratinocyte growth factor or fibroblast growth factor-10 promotes cell proliferation in human endometrial carcinoma cells*. J Clin Endocrinol Metab 2003, 88(2): 773-80.
56. Zang, X.P., Siwak, D., Nguyen, T.X., Tari, A.M., Pento, J.T. *KGF-induced motility of breast cancer cells is dependent on Grb2 and Erk1,2*. Clin Exp Metastasis 2004, 21(5): 437-43.
57. Sharma, G.D., He, J., Bazan, H.E. *p38 and ERK1/2 coordinate cellular migration and proliferation in epithelial wound healing: evidence of cross-talk activation between MAP kinase cascades*. J Biol Chem 2003, 278(24): 21989-97.
58. Zang, X.P., Lerner, M.L., Do, S.V., Brackett, D.J., Pento, J.T. *Keratinocyte growth factor-mediated pattern of gene expression in breast cancer cells*. Cancer Genomics Proteomics 2004, 1(4): 339-44.
59. Skolnik, E.Y., Batzer, A., Li, N. et al. *The function of GRB2 in linking the insulin receptor to Ras signaling pathways*. Science 1993, 260(5116): 1953-5.
60. Warne, P.H., Vician, P.R., Downward, J. *Direct interaction of Ras and the amino-terminal region of Raf-1 in vitro*. Nature 1993, 364(6435): 352-5.
61. Tari, A.M., Hung, M.C., Li, K., Lopez-Berestein, G. *Growth inhibition of breast cancer cells by Grb2 downregulation is correlated with inactivation of mitogen-activated protein kinase in EGFR, but not in ErbB2, cells*. Oncogene 1999, 18(6): 1325-32.
62. Klint, P., Kanda, S., Claesson-Welsh, L. *Shc and a novel 89-kDa component couple to the Grb2-Sos complex in fibroblast growth factor-2-stimulated cells*. J Biol Chem 1995, 270(40): 23337-44.
63. Shono, T., Kanetake, H., Kanda, S. *The role of mitogen-activated protein kinase activation within focal adhesions in chemotaxis toward FGF-2 by murine brain capillary endothelial cells*. Exp Cell Res 2001, 264(2): 275-83.
64. Daly, R.J., Binder, M.D., Sutherland, R.L. *Overexpression of the Grb2 gene in human breast cancer cell lines*. Oncogene 1994, 9(9): 2723-7.
65. Verbeek, B.S., Adriaansen-Slot, S.S., Rijkse, G., Vroom, T.M. *Grb2 overexpression in nuclei and cytoplasm of human breast cells: A histochemical and biochemical study of normal and neoplastic mammary tissue specimens*. J Pathol 1997, 183(2): 195-203.
66. Dickson, C., Spencer-Dene, B., Dillon, C., Fantl, V. *Tyrosine kinase signalling in breast cancer: Fibroblast growth factors and their receptors*. Breast Cancer Res 2000, 2(3): 191-6.
67. Vlodavsky, I., Miao, H.Q., Medalion, B., Danagher, P., Ron, D. *Involvement of heparan sulfate and related molecules in sequestration and growth promoting activity of fibroblast growth factor*. Cancer Metastasis Rev 1996, 15(2): 177-86.
68. Partin, A.W., Schoeniger, J.S., Mohler, J.L., Coffey, D.S. *Fourier analysis of cell motility: Correlation of motility with metastatic potential*. Proc Natl Acad Sci U S A 1989, 86(4): 1254-8.
69. Hiscox, S., Hallett, M.B., Puntis, M.C., Jiang, W.G. *Inhibition of cancer cell motility and invasion by interleukin-12*. Clin Exp Metastasis 1995, 13(5): 396-404.
70. Watanabe, H., Shinozaki, T., Raz, A., and Chigira, M. *Expression of autocrine motility factor receptor in serum- and protein-independent fibrosarcoma cells: implications for autonomy in tumor-cell motility and metastasis*. Int J Cancer 1993, 53(4): 689-95.
71. Peehl, D.M., Rubin, J.S. *Keratinocyte growth factor: an androgen-regulated mediator of stromal-epithelial interactions in the prostate*. World J Urol 1995, 13(5): 312-7.
72. Matsumoto, K., Date, K., Ohmichi, H., Nakamura, T. *Hepatocyte growth factor in lung morphogenesis and tumor invasion: Role as a mediator in epithelium-mesenchyme and tumor-stroma interactions*. Cancer Chemother Pharmacol 1996, 38 Suppl: S42-7.
73. Zang, X-P., Lerner, M.R., Bahr, S.J., Brackett, D.J., Pento, J.T. *A comparison of KGF receptor expression in various types of human cancer*. Cancer Genom Proteom 2006, 3(6): 369-72.
74. Ulich, T.R., Yi, E.S., Cardiff, R. et al. *Keratinocyte growth factor is a growth factor for mammary epithelium in vivo. The mammary epithelium of lactating rats is resistant to the proliferative action of keratinocyte growth factor*. Am J Pathol 1994, 144(5): 862-8.
75. Kitsberg, D.I., Leder, P. *Keratinocyte growth factor induces mammary and prostatic hyperplasia and mammary adenocarcinoma in transgenic mice*. Oncogene 1996, 13(12): 2507-15.
76. Koos, R.D., Banks, P.K., Inkster, S.E., Yue, W., Brodie, A.M. *Detection of aromatase and keratinocyte growth factor expression in breast tumors using reverse transcription-polymerase chain reaction*. J Steroid Biochem Mol Biol 1993, 45(4): 217-25.

77. Bansal, G.S., Cox, H.C., Marsh, S. et al. *Expression of keratinocyte growth factor and its receptor in human breast cancer*. Br J Cancer 1997, 75(11): 1567-74.
78. Lochter, A., Galosy, S., Muschler, J., Freedman, N., Werb, Z., Bissell, M.J. *Matrix metalloproteinase stromelysin-1 triggers a cascade of molecular alterations that leads to stable epithelial-to-mesenchymal conversion and a premalignant phenotype in mammary epithelial cells*. J Cell Biol 1997, 139(7): 1861-72.
79. Zang, X., Learner, M.L., Brackett, D.J., Pento, J.T. *KGF-induced gene expression in MCF-7 cells using cDNA expression arrays*. Breast Cancer Res Treat [23rd Annual San Antonio Breast Cancer (December 6-9, San Antonio, Texas, 2000)] 2000, 64(1): 110.
80. Zang, X.P., Pento, J.T. *Keratinocyte growth factor-induced motility of breast cancer cells*. Clin Exp Metastasis 2000, 18(7): 573-80.
81. Zang, X.P., Bullen, E.C., Manjeshwar, S., Jupe, E.R., Howard, E.W., Pento, J.T. *Enhanced motility of KGF-transfected breast cancer cells*. Anticancer Res 2006, 26(2A): 961-6.
82. Cheng, T.P. *Minipodia, novel structures for extension of the lamella: A high-spatial-resolution video microscopic study*. Exp Cell Res 1992, 203(1): 25-31.
83. an Larebeke, N.A., Bracke, M.E., Mareel, M.M. *Invasive epithelial cells show more fast plasma membrane movements than related or parental non-invasive cells*. Cytometry 1992, 13(1): 9-14.
84. Rajah, T.T., Rambo, D.J., Dmytryk, J.J., Pento, J.T. *Influence of antiestrogens on NIH-3T3-fibroblast-induced motility of breast cancer cells*. Chemotherapy 2001, 47(1): 56-69.
85. Hishikawa, Y., Tamaru, N., Ejima, K., Hayashi, T., Koji, T. *Expression of keratinocyte growth factor and its receptor in human breast cancer: Its inhibitory role in the induction of apoptosis possibly through the overexpression of Bcl-2*. Arch Histol Cytol 2004, 67(5): 455-64.
86. Zang, X.P., Pento, J.T., Tari, A.M. *Wilms' tumor 1 protein and focal adhesion kinase mediate keratinocyte growth factor signaling in breast cancer cells*. Anticancer Res 2008, 28(1A): 133-7.
87. Miyoshi, Y., Ando, A., Egawa, C. et al. *High expression of Wilms' tumor suppressor gene predicts poor prognosis in breast cancer patients*. Clin Cancer Res 2002, 8(5): 1167-71.
88. Schlaepfer, D.D., Hanks, S.K., Hunter, T., van der Geer, P. *Integrin-mediated signal transduction linked to Ras pathway by GRB2 binding to focal adhesion kinase*. Nature 1994, 372(6508): 786-91.
89. Tuna, M., Chavez-Reyes, A., Tari, A.M. *HER2/neu increases the expression of Wilms' Tumor 1 (WT1) protein to stimulate S-phase proliferation and inhibit apoptosis in breast cancer cells*. Oncogene 2005, 24(9): 1648-52.
90. Aznavoorian, S., Murphy, A.N., Stetler-Stevenson, W.G., Liotta, L.A. *Molecular aspects of tumor cell invasion and metastasis*. Cancer 1993, 71(4): 1368-83.
91. Tamaru, N., Hishikawa, Y., Ejima, K. et al. *Estrogen receptor-associated expression of keratinocyte growth factor and its possible role in the inhibition of apoptosis in human breast cancer*. Lab Invest 2004, 84(11): 1460-71.
92. Zang, X.P., Lerner, M.R., Dunn, S.T., Brackett, D.J., Pento, J.T. *Antisense KGF oligonucleotide inhibition of KGF-induced motility in breast cancer cells*. Anticancer Res 2003, 23(6C): 4913-9.
93. Zang, X.P., Pento, J.T. *siRNA inhibition of ER-alpha expression reduces KGF-induced proliferation of breast cancer cells*. Anticancer Res 2008, 28(5A): 2733-5.
94. Leung, H.Y., Mehta, P., Gray, L.B., Collins, A.T., Robson, C.N., Neal, D.E. *Keratinocyte growth factor expression in hormone insensitive prostate cancer*. Oncogene 1997, 15(9): 1115-20.
95. McCarvey, T.W., Stearns, M.E. *Keratinocyte growth factor and receptor mRNA expression in benign and malignant human prostate*. Exp Mol Pathol 1995, 63(1): 52-62.
96. Planz, B., Oltean, H., Deix, T., Kirley, S.D., Wang, Q.F., McDougal, W.S., Marberger, M. *Effect of keratinocyte growth factor and activin on cell growth in the human prostatic cancer cell line LNCaP*. World J Urol 2004, 22(2): 140-4.
97. De Bellis, A., Crescioli, C., Grappone, C., Milani, S., Ghiandi, P., Forti, G., Serio, M. *Expression and cellular localization of keratinocyte growth factor and its receptor in human hyperplastic prostate tissue*. J Clin Endocrinol Metab 1998, 83(6): 2186-91.
98. Kaminski, A., Hahne, J.C., Haddouti el-M., Florin, A., Wellmann, A., Wernert, N. *Tumour-stroma interactions between metastatic prostate cancer cells and fibroblasts*. Int J Mol Med 2006, 18(5): 941-50.
99. Planz, B., Aretz, H.T., Wang, Q., Tabatabaei, S., Kirley, S.D., Lin, C.W., McDougal, W.S. *Immunolocalization of the keratinocyte growth factor in benign and neoplastic human prostate and its relation to androgen receptor*. Prostate 1999, 41(4): 233-42.
100. Yan, G., Fukabori, Y., McBride, G., Nikolopoulos, S., McKeenan, W.L. *Exon switching and activation of stromal and embryonic fibroblast growth factor (FGF)-FGF receptor genes in prostate epithelial cells accompany stromal independence and malignancy*. Mol Cell Biol 1993, 13(8): 4513-22.
101. Foster, B.A., Kaplan, P.J., Greenberg, N.M. *Characterization of the FGF axis and identification of a novel FGFR3iic isoform during prostate cancer progression in the TRAMP model*. Prostate Cancer Prostatic Dis 1999, 2(2): 76-82.
102. Feng, S., Wang, F., Matsubara, A., Kan, M., McKeenan, W.L. *Fibroblast growth factor receptor 2 limits and receptor 1 accelerates tumorigenicity of prostate epithelial cells*. Cancer Res 1997, 57(23): 5369-78.
103. Ropiquet, F., Giri, D., Lamb, D.J., Ittmann, M. *FGF7 and FGF2 are increased in benign prostatic hyperplasia and are associated with increased proliferation*. J Urol 1999, 162(2): 595-9.
104. Ropiquet, F., Huguenin, S., Villette, J.M. et al. *FGF7/KGF triggers cell transformation and invasion on immortalised human prostatic epithelial PNT1A cells*. Int J Cancer 1999, 82(2): 237-43.
105. Cussenot, O. [Growth factors and prostatic tumors.] Ann Endocrinol (Paris) 1997, 58(5): 370-80.
106. Otte, J.M., Schmitz, F., Banasiewicz, T., Drews, M., Fölsch, U.R., Herzig, K.H. *Expression of keratinocyte growth factor and its receptor in colorectal cancer*. Eur J Clin Invest 2000, 30(3): 222-9.
107. Dignass, A.U., Tsunekawa, S., Podolsky, D.K. *Fibroblast growth factors modulate intestinal epithelial cell growth and migration*. Gastroenterology 1994, 106(5): 1254-62.
108. Jonas, C.R., Gu, L.H., Nkabyo, Y.S. et al. *Glutamine and KGF each regulate extracellular thiol/disulfide redox and enhance proliferation in Caco-2 cells*. Am J Physiol Regul Integr Comp Physiol 2003, 285(6): R1421-9.
109. Yoshino, M., Ishiwata, T., Watanabe, M., Komine, O., Shibuya, T., Tokunaga, A., Naito, Z. *Keratinocyte growth factor receptor expression in normal colorectal epithelial cells and differentiated type of colorectal cancer*. Oncol Rep 2005, 13(2): 247-52.
110. Moll, R., Löwe, A., Laufer, J., Franke, W.W. *Cytokeratin 20 in human carcinomas. A new histodiagnostic marker detected by monoclonal antibodies*. Am J Pathol 1992, 140(2): 427-447.
111. Watanabe, M., Ishiwata, T., Nishigai, K., Moriyama, Y., Asano, G. *Overexpression of keratinocyte growth factor in cancer cells and enterochromaffin cells in human colorectal cancer*. Pathol Int 2000, 50(5): 363-72.
112. Fehrenbach, H., Kasper, M., Koslowski, R., Pan, T., Schuh, D., Müller, M., Mason, R.J. *Alveolar epithelial type II cell apoptosis in vivo during resolu-*

- tion of keratinocyte growth factor-induced hyperplasia in the rat. *Histochem Cell Biol* 2000, 114(1): 49-61.
113. Yamayoshi, T., Nagayasu, T., Matsumoto, K., Abo, T., Hishikawa, Y., Koji, T. *Expression of keratinocyte growth factor/fibroblast growth factor-7 and its receptor in human lung cancer: Correlation with tumour proliferative activity and patient prognosis.* *J Pathol* 2004, 204(1): 110-8.
 114. Baumann, M., Krause, M., Zips, D., Petersen, C., Dittmann, K., Dörr, W., Rodemann, H.P. *Molecular targeting in radiotherapy of lung cancer.* *Lung Cancer* 2004, 45 Suppl 2: S187-97.
 115. Steele, I.A., Edmondson, R.J., Bulmer, J.N., Bolger, B.S., Leung, H.Y., Davies, B.R. *Induction of FGF receptor 2-IIIb expression and response to its ligands in epithelial ovarian cancer.* *Oncogene* 2001, 20(41): 5878-87.
 116. Yasuhara, T., Okamoto, A., Kitagawa, T. et al. *FGF7-like gene is associated with pericentric inversion of chromosome 9, and FGF7 is involved in the development of ovarian cancer.* *Int J Oncol* 2005, 26(5): 1209-16.
 117. Parrott, J.A., Kim, G., Mosher, R., Skinner, M.K. *Expression and action of keratinocyte growth factor (KGF) in normal ovarian surface epithelium and ovarian cancer.* *Mol Cell Endocrinol* 2000, 167(1-22): 77-87.
 118. Kurban, G., Ishiwata, T., Kudo, M., Yokoyama, M., Sugisaki, Y., Naito, Z. *Expression of keratinocyte growth factor receptor (KGFR/FGFR2 IIIb) in human uterine cervical cancer.* *Oncol Rep* 2004, 11(5): 987-91.
 119. Visco, V., Carico, E., Marchese, C., Torrissi, M.R., Frati, L., Vecchione, A., Muraro, R. *Expression of keratinocyte growth factor receptor compared with that of epidermal growth factor receptor and erbB-2 in endometrial adenocarcinoma.* *Int J Oncol* 1999, 15(3): 431-5.
 120. Yeh, J.J. *Prognostic signature for pancreatic cancer: are we close?* *Future Oncol* 2009, 5(3): 313-21.
 121. Siddiqi, I., Funatomi, H., Kobrin, M.S., Friess, H., Büchler, M.W., Korc, M. *Increased expression of keratinocyte growth factor in human pancreatic cancer.* *Biochem Biophys Res Commun* 1995, 215(1): 309-15.
 122. Otte, J.M., Schwenger, M., Brunke, G. et al. *Expression of hepatocyte growth factor, keratinocyte growth factor and their receptors in experimental chronic pancreatitis.* *Eur J Clin Invest* 2001, 31(10): 865-75.
 123. Ishiwata, T., Friess, H., Büchler, M.W., Lopez, M.E., Korc, M. *Characterization of keratinocyte growth factor and receptor expression in human pancreatic cancer.* *Am J Pathol* 1998, 153(1): 213-22.
 124. Hasegawa, Y., Takada, M., Yamamoto, M., Saitoh, Y. *The gradient of basic fibroblast growth factor concentration in human pancreatic cancer cell invasion.* *Biochem Biophys Res Commun* 1994, 200(3): 1435-9.
 125. Kornmann, M., Ishiwata, T., Matsuda, K. et al. *IIIc isoform of fibroblast growth factor receptor 1 is overexpressed in human pancreatic cancer and enhances tumorigenicity of hamster ductal cells.* *Gastroenterology* 2002, 123(1): 301-13.
 126. Wagner, M., Lopez, M.E., Cahn, M., Korc, M. *Suppression of fibroblast growth factor receptor signaling inhibits pancreatic cancer growth in vitro and in vivo.* *Gastroenterology* 1998, 114(4): 798-807.
 127. Zang, X.P., Lerner, M.L., Brackett, D.J., Pento, J.T. *Influence of KGF on the progression of pancreatic cancer.* *Anticancer Res* 2009, 29(8): 3417-20.
 128. Kiehne, K., Otte, J.M., Folsch, U.R., Herzig, K.H. *Growth factors in development and diseases of the exocrine pancreas.* *Pancreatol* 2001, 1(1): 15-23.
 129. Vickers, S.M., Huang, Z.Q., MacMillan-Crow, L., Greendorfer, J.S., Thompson, J.A. *Ligand activation of alternatively spliced fibroblast growth factor receptor-1 modulates pancreatic adenocarcinoma cell malignancy.* *J Gastrointest Surg* 2002, 6(4): 546-53.
 130. Hattori, Y., Odagiri, H., Nakatani, H. et al. *K-sam, an amplified gene in stomach cancer, is a member of the heparin-binding growth factor receptor genes.* *Proc Natl Acad Sci U S A* 1990, 87(15): 5983-7.
 131. Shin, E.Y., Lee, B.H., Yang, J.H. et al. *Up-regulation and co-expression of fibroblast growth factor receptors in human gastric cancer.* *J Cancer Res Clin Oncol* 2000, 126(9): 519-28.
 132. Nakazawa, K., Yashiro, M., Hirakawa, K. *Keratinocyte growth factor produced by gastric fibroblasts specifically stimulates proliferation of cancer cells from scirrhous gastric carcinoma.* *Cancer Res* 2003, 63(24): 8848-52.
 133. Matsunobu, T., Ishiwata, T., Yoshino, M. et al. *Expression of keratinocyte growth factor receptor correlates with expansive growth and early stage of gastric cancer.* *Int J Oncol* 2006, 28(2): 307-14.
 134. Zacharski, L.R., Ornstein, D.L. *Heparin and cancer.* *Thromb Haemost* 1998, 80(1): 10-23.
 135. Bonneh-Barkay, D., Shlissel, M., Berman, B. et al. *Identification of glypican as a dual modulator of the biological activity of fibroblast growth factors.* *J Biol Chem* 1997, 272(19): 12415-21.
 136. Ron, D., Bottaro, D.P., Finch, P.W., Morris, D., Rubin, J.S., Aaronson, S.A. *Expression of biologically active recombinant keratinocyte growth factor. Structure/function analysis of amino-terminal truncation mutants.* *J Biol Chem* 1993, 268(4): 2984-8.
 137. Gould, M.K., Dembitzer, A.D., Doyle, R.L., Hastie, T.J., Garber, A.M. *Low-molecular-weight heparins compared with unfractionated heparin for treatment of acute deep venous thrombosis. A meta-analysis of randomized, controlled trials.* *Ann Intern Med* 1999, 130(10): 800-9.
 138. Gould, M.K., Dembitzer, A.D., Sanders, G.D., Garber, A.M. *Low-molecular-weight heparins compared with unfractionated heparin for treatment of acute deep venous thrombosis. A cost-effectiveness analysis.* *Ann Intern Med* 1999, 130(10): 789-99.
 139. Zang, X.P., Nguyen, T.N., Pento, J.T. *Specific and non-specific KGF inhibition of KGF-induced breast cancer cell motility.* *Anticancer Res* 2002, 22(5): 2539-45.
 140. Galzie, Z., Kinsella, A.R., Smith, J.A. *Fibroblast growth factors and their receptors.* *Biochem Cell Biol* 1997, 75(6): 669-85.
 141. Liu, D., Shriver, Z., Qi, Y., Venkataraman, G., Sasisekharan, R. *Dynamic regulation of tumor growth and metastasis by heparan sulfate glycosaminoglycans.* *Semin Thromb Hemost* 2002, 28(1): 67-78.
 142. Liu, D., Shriver, Z., Venkataraman, G., El Shabrawi, Y., Sasisekharan, R. *Tumor cell surface heparan sulfate as cryptic promoters or inhibitors of tumor growth and metastasis.* *Proc Natl Acad Sci U S A* 2002, 99(2): 568-73.
 143. Sasisekharan, R., Shriver, Z., Venkataraman, G., Narayanasami, U. *Roles of heparan-sulphate glycosaminoglycans in cancer.* *Nat Rev Cancer* 2002, 2(7): 521-8.
 144. Eisenberger, M.A., Reyno, L.M., Jodrell, D.I. et al. *Suramin, an active drug for prostate cancer: Interim observations in a phase I trial.* *J Natl Cancer Inst* 1993, 85(8): 611-21.
 145. Motzer, R.J., Nanus, D.M., O'Moore, P. et al. *Phase II trial of suramin in patients with advanced renal cell carcinoma: Treatment results, pharmacokinetics, and tumor growth factor expression.* *Cancer Res* 1992, 52(20): 5775-9.
 146. Walther, M.M., Figg, W.D., Linehan, W.M. *Intravesical suramin: A novel agent for the treatment of superficial transitional-cell carcinoma of the bladder.* *World J Urol* 1996, 14 Suppl 1: S8-11.
 147. Zhang, Y., Song, S., Yang, F., Au, J.L., Wientjes, M.G. *Nontoxic doses of suramin enhance activity of doxorubicin in prostate tumors.* *J Pharmacol Exp Ther* 2001, 299(2): 426-33.
 148. Rotolo, S., Ceccarelli, S., Romano, F., Frati, L., Marchese, C., Angeloni, A. *Silencing of keratinocyte growth factor receptor restores 5-fluorouracil and tamoxifen efficacy on responsive cancer cells.* *PLoS One* 2008, 3(6): e2528.

149. Kudchadkar, R., Gonzalez, R., Lewis, K.D. *PI-88: A novel inhibitor of angiogenesis*. *Expert Opin Investig Drugs* 2008, 17(11): 1769-76.
 150. Marchiani, S., Bonaccorsi, L., Ferruzzi, P. et al. *The vitamin D analogue BXL-628 inhibits growth factor-stimulated proliferation and invasion of DU145 prostate cancer cells*. *J Cancer Res Clin Oncol* 2006, 132(6): 408-16.
 151. Traxler, P., Bold, G., Buchdunger, E. et al. *Tyrosine kinase inhibitors: From rational design to clinical trials*. *Med Res Rev* 2001, 21(6): 499-512.
 152. Zwick, E., Bange, J., Ullrich, A. *Receptor tyrosine kinases as targets for anticancer drugs*. *Trends Mol Med* 2002, 8(1):17-23.
 153. Klijn, J.G., Berns, P.M., Schmitz, P.I., Foekens, J.A. *The clinical significance of epidermal growth factor receptor (EGF-R) in human breast cancer: A review on 5232 patients*. *Endocr Rev* 1992, 13(1): 3-17.
 154. Woodburn, J.R. *The epidermal growth factor receptor and its inhibition in cancer therapy*. *Pharmacol Ther* 1999, 82(2-3): 241-250.
 155. Sainsbury, J.R., Farndon, J.R., Needham, G.K., Malcolm, A.J., Harris, A.L. *Epidermal-growth-factor receptor status as predictor of early recurrence of and death from breast cancer*. *Lancet* 1987, 1(8547): 1398-402.
 156. Druker, B.J. *STI571 (Gleevec) as a paradigm for cancer therapy*. *Trends Mol Med* 2002, 8(4 Suppl): S14-8.
 157. Janmaat, M.L., and Giaccone, G. *The epidermal growth factor receptor pathway and its inhibition as anticancer therapy*. *Drugs Today (Barc)* 2003, 39 Suppl C: 61-80.
 158. Nguyen, T., Nguyen, T.N., Zang, X.P., and Pento, J.T. 2003. KGF signal transduction pathways involved in breast cancer cell metastasis. *OUHSC Great Symposium*:25.
 159. García-Echeverría, C., Traxler, P., Evans, D.B. *ATP site-directed competitive and irreversible inhibitors of protein kinases*. *Med Res Rev* 2000, 20(1): 28-57.
 160. Toledo, L.M., Lydon, N.B., Elbaum, D. *The structure-based design of ATP-site directed protein kinase inhibitors*. *Curr Med Chem* 1999, 6(9): 775-805.
 161. Hackett, J., Xiao, Z., Zang, X.P. *Development of keratinocyte growth factor receptor tyrosine kinase inhibitors for the treatment of cancer*. *Anticancer Res* 2007, 27(6B): 3801-6.
-